

The reproducibility and comparability of tests of the peripheral chemoreflex: comparing the transient hypoxic ventilatory drive test and the single-breath carbon dioxide response test in healthy subjects

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Abstract. Both the transient hypoxic ventilatory drive test and the single-breath carbon dioxide (CO₂) response test have been used to assess peripheral chemoreflex sensitivity. We tested their comparability in 14 healthy adults (10 men, aged 31–73 years, mean 55.4 years). The within-subject reproducibility of both tests was also assessed ($n=7$ for each). The mean transient hypoxic ventilatory response was $0.287 \pm 0.059 \text{ l min}^{-1} (\% \text{SaO}_2)^{-1}$ (mean \pm SEM, range 0.018–0.718) and single-breath CO₂ response was $0.276 \pm 0.041 \text{ l min}^{-1} \text{ T}^{-1}$ (range 0.081–0.501). Both tests were reproducible with a mean coefficient of variation of 20.1% and 17.7%, respectively. There was, however, no significant correlation between the results of the transient hypoxic and single-breath CO₂ tests when data were compared by linear regression analysis ($r=0.23$, $P=0.43$), suggesting that separate pathways of the peripheral chemoreflex existed for hypoxia and hypercapnia, respectively, and that these tests were specific for each. The authors conclude that these tests are reproducible but need to be used in combination for an adequate assessment of the peripheral chemoreflex.

Keywords. Peripheral chemoreflex, single-breath carbon dioxide, transient hypoxia, ventilatory responses.

Introduction

Peripheral chemoreceptor responsiveness is generally assessed by hypoxic testing, and three major techniques are available. These are the steady state [1], progressive [2,3] and transient hypoxic methods [4]. They all appear to correlate well with each other [5,6]. However, persistent hypoxia especially with the first two methods may result in the depression of ventilation owing to direct central effects. There are

also the potential risks of hypoxic exposure with these tests. McClean *et al.* [7] reported the use of the single-breath carbon dioxide response test as an alternative method of assessing peripheral chemoreflex and highlighted the relative safety of the test. Abnormalities in regulation of ventilation may exist in certain lung and heart disorders [8–14], and the magnitude of respiratory drive may contribute to the degree of breathlessness. Because of the ease of performing the transient hypoxic and the single-breath carbon dioxide response tests, they are ideally suited as clinical tests of the peripheral chemoreflex. They may be useful in clinical practice to assess patients with chronic lung disease, asthma, heart diseases and unexplained dyspnoea. We are, however, unaware of any study in which the relation between the two tests is assessed. We therefore proceeded to examine the reproducibility of and the relationship between the tests.

Patients and methods

Fourteen healthy subjects, 10 men and four women, participated in this study. They were 31–73 years of age and were either staff members of the Royal Brompton National Heart & Lung Institute or friends of patients of the affiliated Royal Brompton National Heart & Lung Hospital. None had respiratory symptoms and all except two were non-smokers. All were told to avoid caffeinated products on the morning of the tests. Subject characteristics, including the results of spirometry tests, are summarized in Table 1.

All 14 subjects underwent both the transient hypoxic ventilatory drive test and the single-breath carbon dioxide response test on separate occasions. In addition, the transient hypoxic ventilatory drive test and the single-breath carbon dioxide response test were each repeated once within a month in seven subjects for the assessment of within-subject reproducibility. The study had been approved by the local ethics committee.

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Table 1. Subject characteristics including results of spirometry tests

| Number of subjects | Age (years) | Height (cm) | Weight (kg) | Body surface Area (m ²) | FEV ₁ (% predicted) | FVC (% predicted) |
|--------------------|-----------------------|--------------------------|-----------------------|-------------------------------------|--------------------------------|-------------------------|
| 14 (10 men) | 55.4 ± 3.2 (31–73) | 168.6 ± 2.3 (156–180) | 71.8 ± 3.8 (45–93) | 1.87 ± 0.05 (1.48–2.14) | 104.4 ± 4.4 (82–140) | 111.8 ± 4.7 (91–144) |

FEV₁, forced expiratory volume in 1 s; FVC, forced vital capacity. Body surface area is obtained from a standard height–weight–body surface area nomogram [15]. Values expressed as means ± SEM (range).

Transient hypoxic ventilatory drive test

The method of performing the transient hypoxic test was described by Edelman *et al.* [4]. This was performed while subjects were seated and after a period of quiet breathing. They were encouraged to relax in a quiet environment and were allowed to read. Each subject wore a noseclip and breathed through a pneumatic respiratory valve (Innovision, Odense, Denmark), which separated the expirate from the inspirate. The inspirate port was further connected to a T-valve placed behind the subject, and depending on the position of the T-valve, the subject breathed either room air or pure nitrogen from a 4-L reservoir bag. This bag was quietly refilled from a gas cylinder containing pure nitrogen with a valve mechanism (Intersurgical Complete Respiratory Systems, Wokingham, UK), which prevented overfilling and pressure from building up in the bag. Minute ventilation was measured breath by breath using a heated pneumotachograph by integrating the flow over one whole expiration and dividing by the duration of the breath. This was done on line. O₂ and CO₂ at the mouth were monitored continuously by mass spectrometry (Amis 2000, Innovision). The pneumotachometer and mass spectrometer were calibrated before each test. Arterial oxygen saturation was measured using a pulse oximeter (Model N-200E, Nellcor, Hayward, CA, USA) set at fast mode with a response time of 2–3 s and a lightweight ear-probe clipped gently on the subject's right ear lobe.

After the subject had breathed room air for several minutes, surreptitiously and without the subject's knowledge, the T-valve was turned during the expiratory phase of one breath so that pure nitrogen was inhaled for the next 2–8 breaths. This was repeated 10–15 times so as to provide a wide range of arterial oxygen saturations from 70% to 100%. End-tidal O₂ was monitored during the testing for safety reasons to prevent extreme hypoxia. Each transient was preceded by a period of air breathing, during which time oxygen saturation and end-tidal CO₂ were allowed to return to the subject's baseline. The average of the two largest consecutive breaths that gave the highest ventilation after the hypoxic stimulus was used to calculate maximal minute ventilation. A two-breath period was used for calculation of highest ventilation rather than a single breath to reduce data variability (unpublished observations and also noted by Edelman *et al.* [4]).

This value was plotted against the lowest arterial oxygen desaturation reached for that period of nitrogen inhalation. The ventilatory response to transient hypoxia was expressed as the slope that related ventilation and arterial oxygen saturation, calculated by least-squares linear regression analysis, in terms of litres per minute per per cent O₂ saturation [L min⁻¹ (%SaO₂)⁻¹].

Single-breath carbon dioxide response test

The method used was similar to that described by McClean *et al.* [7]. The apparatus including the T-valve was the same as for the transient hypoxic ventilatory drive test above. A smaller 2 L reservoir bag was used, and this was quietly refilled after each inhalation with a gas mixture containing 13% CO₂ in air instead of pure nitrogen. After a period of quiet breathing, the T-valve was again turned in a surreptitious manner during the expiratory phase of one breath such that the subject next inhaled a single breath of gas mixture high in CO₂ content. On average, 10 single breaths of CO₂ were administered at approximately 2 min intervals. As before, minute ventilation was measured breath by breath using a heated pneumotachograph and continuous monitoring of CO₂ was done at the mouth by mass spectrometry.

The mean of the minute ventilation of the preceding five breaths before the stimulus CO₂ breath was calculated and taken as the control ventilation [$\dot{V}(C)$]. The mean end-tidal fraction of CO₂ of these breaths was also calculated and taken as the control end-tidal fraction of CO₂ [$F_{ETCO_2}(C)$]. The response ventilation after the stimulus CO₂ breath [$\dot{V}(S)$] was calculated by averaging the two largest consecutive breaths but, unlike hypoxic testing, breaths during the first 20 s after the stimulus breath were excluded. This is because breaths beyond this time limit were assumed to be affected by changes in central chemoreceptor drive [7]. The end-tidal CO₂ concentration after the stimulus breath was considered the stimulus end-tidal fraction of CO₂ [$F_{ETCO_2}(S)$].

The single-breath CO₂ response was calculated using the parameters above as follows:

$$= \frac{\dot{V}(S) - \dot{V}(C)}{[F_{ETCO_2}(S) - F_{ETCO_2}(C)] \times (P_B - 47)}$$

Table 2. Results of transient hypoxic ventilatory drive and single-breath CO₂

| Subject | Age (years) | Sex | Hypoxic ventilatory response [L min ⁻¹ (%SaO ₂) ⁻¹] | Single-breath CO ₂ ventilatory response (L min ⁻¹ T ⁻¹) |
|---------|-------------|-----|--|---|
| 1 | 50 | F | 0.182 | 0.390 |
| 2 | 64 | F | 0.082 | 0.110 |
| 3 | 64 | M | 0.615 | 0.268 |
| 4 | 56 | M | 0.183 | 0.092 |
| 5 | 58 | F | 0.186 | 0.081 |
| 6 | 31 | M | 0.236 | 0.495 |
| 7 | 69 | M | 0.222 | 0.446 |
| 8 | 42 | M | 0.299 | 0.182 |
| 9 | 64 | M | 0.265 | 0.232 |
| 10 | 41 | M | 0.168 | 0.501 |
| 11 | 45 | M | 0.018 | 0.270 |
| 12 | 58 | M | 0.718 | 0.309 |
| 13 | 61 | F | 0.157 | 0.083 |
| 14 | 73 | M | 0.687 | 0.406 |
| Mean | 55.4 | | 0.287 | 0.276 |
| SEM | 3.2 | | 0.059 | 0.041 |

where P_B is the atmospheric pressure in torr and 47 is the saturated water vapour pressure in torr. The mean of 10 responses was considered to be the subject's single-breath CO₂ response and expressed in litres per minute per torr (L min⁻¹ T⁻¹).

Statistical analysis

Studies of reproducibility within subjects and the relation between the transient hypoxic and single-breath CO₂ tests were assessed by linear regression analysis. The coefficients of variation of the tests were also calculated. $P < 0.05$ was considered significant.

Results

The results of the transient hypoxic and single-breath CO₂ response tests are summarized in Table 2. The

mean transient hypoxic ventilatory response was 0.287 ± 0.059 L min⁻¹ (%SaO₂)⁻¹ and the mean single-breath CO₂ response 0.276 ± 0.041 L min⁻¹ T⁻¹. The transient hypoxic responses correlated significantly with height ($r = 0.57$, $P = 0.03$) and body surface area ($r = 0.53$, $P = 0.049$), but not with weight ($r = 0.41$, $P = 0.14$). There was an association between single-breath CO₂ responses and height, although this did not achieve statistical significance ($r = 0.51$, $P = 0.06$). No correlation was found between single-breath CO₂ responses and weight ($r = 0.3$, $P = 0.75$) or body surface area ($r = 0.35$, $P = 0.21$). Neither transient hypoxic nor single-breath CO₂ responses correlated with age or spirometric measurements.

The reproducibility between two tests taken on different days is shown in Tables 3 and 4. The coefficient of variation calculated from the results of two transient hypoxic tests in seven subjects varied from 9.4% to 33% with a mean of $20.1\% \pm 3.3\%$. Table 3 also shows the correlation coefficients obtained from the plots relating \dot{V}_E and arterial oxygen desaturation, with a mean r of 0.75 for the first test and of 0.74 for the second. Ventilatory responses were linear in relation to arterial oxygen desaturation in all tests. Figure 1 shows a graphical representation of the reproducibility of the transient hypoxic test ($r = 0.96$, $P < 0.001$).

The mean coefficient of variation obtained from the two single-breath CO₂ tests in seven subjects was $17.7\% \pm 6.7\%$ with a range of 3.9–56%. The high coefficient of variation of 56% seen in subject 2 may be the result of a low mean single-breath CO₂ response of this subject since the coefficient of variation is a measure of variability in relation to the magnitude of the mean. Figure 2 shows the results of the two single-breath CO₂ tests plotted graphically ($r = 0.84$, $P = 0.02$).

There was no significant correlation between the transient hypoxic and single-breath CO₂ responses, as shown in Fig. 3 ($r = 0.23$, $P = 0.43$).

Table 3. Reproducibility of transient hypoxic ventilatory drive test in seven subjects

| Subject | Response 1 | | Response 2 | | Coefficient of variation (%) |
|---------|---|-------|---|-------|------------------------------|
| | [L min ⁻¹ (%SaO ₂) ⁻¹] | r_1 | [L min ⁻¹ (%SaO ₂) ⁻¹] | r_2 | |
| 4 | 0.183 | 0.93 | 0.138 | 0.50 | 19.8 |
| 6 | 0.237 | 0.58 | 0.172 | 0.79 | 22.5 |
| 7 | 0.222 | 0.74 | 0.148 | 0.66 | 28.3 |
| 8 | 0.299 | 0.79 | 0.481 | 0.77 | 33.0 |
| 10 | 0.168 | 0.66 | 0.195 | 0.61 | 10.5 |
| 12 | 0.718 | 0.82 | 0.820 | 0.91 | 9.4 |
| 14 | 0.687 | 0.89 | 0.894 | 0.84 | 18.5 |
| Mean | 0.359 | 0.77 | 0.407 | 0.73 | 20.1 |
| SEM | 0.090 | 0.05 | 0.125 | 0.05 | 3.3 |

r_1 and r_2 are the correlation coefficients, obtained by linear regression analysis from the plot relating \dot{V}_E and arterial oxygen desaturation, during the first and second transient hypoxic ventilatory drive test, respectively.

Table 4. Reproducibility of single-breath CO₂ response test in seven subjects

| Subject | Response 1 (L min ⁻¹ T ⁻¹) | Response 2 (L min ⁻¹ T ⁻¹) | Coefficient of variation (%) |
|---------|---|---|------------------------------|
| 1 | 0.390 | 0.369 | 3.9 |
| 2 | 0.110 | 0.255 | 56.2 |
| 6 | 0.495 | 0.412 | 12.9 |
| 7 | 0.446 | 0.410 | 5.9 |
| 8 | 0.182 | 0.233 | 17.4 |
| 9 | 0.232 | 0.271 | 11.0 |
| 10 | 0.501 | 0.635 | 16.7 |
| Mean | 0.337 | 0.369 | 17.7 |
| SEM | 0.060 | 0.052 | 6.7 |

Discussion

The results indicate that both tests are reproducible. We are not aware of any previous study of reproducibility of the transient hypoxic ventilatory drive test, although one study using the progressive hypoxic test based on the Rebeck & Campbell technique [3] showed a mean coefficient of variation of 23.1% (range 2.3–48.7%) [6]. This is similar to the mean coefficient of variation of 20.1% seen in our study. The mean coefficient of variation for the single-breath CO₂ response test was 17.7% in our study, and this is again compatible with the mean of 25% obtained by McClean *et al.* [7] with the single-breath CO₂ test.

We found that the transient hypoxic ventilatory responses correlated with height but not with age, compatible with the findings of Hirshman *et al.* [16] using the progressive hypoxic method. There was also significant correlation with body surface area. However, we were not able to demonstrate any correlation with weight, as they did, or with spirometric results. The reason for the lack of correlation with weight in our study is unclear. There appears to be no significant correlation between

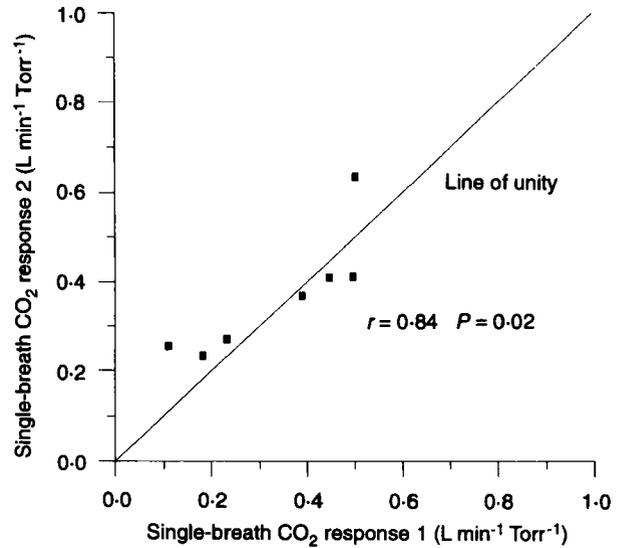


Figure 2. Reproducibility of the single-breath CO₂ response test.

single-breath CO₂ responses and age, height, weight, body surface area or spirometric results. The significance of this observation is not known but may point towards the divergence of the two responses.

We reasoned that if the transient hypoxic ventilatory drive and single-breath CO₂ response tests are assessments of the peripheral chemoreflex, it is likely that there is a significant correlation between the two tests. Our study, however, indicates that there is no correlation between the transient hypoxic and the single-breath CO₂ response tests. It has previously been shown that there is a positive correlation between progressive hypoxic response and CO₂ response by the *rebreathing technique* [17], but it must be borne in mind that the latter is a test of central chemoreceptors [18]. Furthermore, there appears to be no correlation between the single-breath CO₂

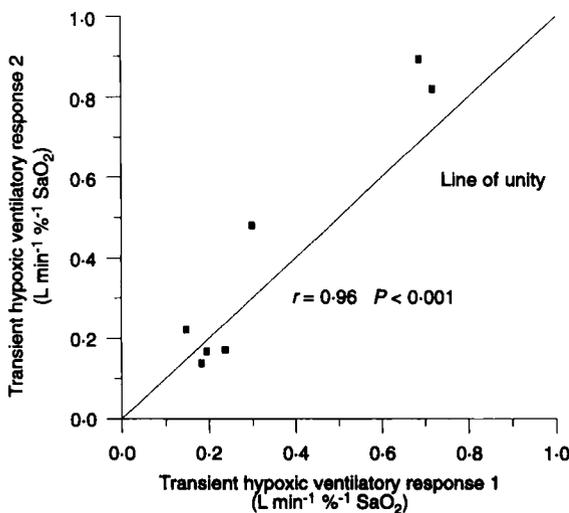


Figure 1. Reproducibility of the transient hypoxic ventilatory drive test.

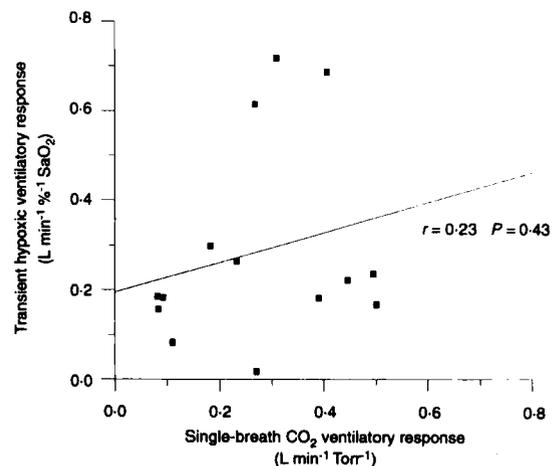


Figure 3. Scatter plot showing the absence of a significant correlation between the transient hypoxic and single-breath CO₂ ventilatory responses.

response and rebreathing CO₂ response [7]. It has also been demonstrated that nitrendipine, a dihydropyridine Ca²⁺-channel blocker, inhibited the release of dopamine, a neurotransmitter in the carotid body chemoreceptors, by hypoxia but not by hypercapnia [19]. This suggests that, although hypoxic and peripheral hypercapnic responses are mediated by the carotid body, the pathways are partially separate, which may account for our observation. When P_{O₂} decreases, Ca²⁺ channels are activated with Ca²⁺ entry, causing the release of neurotransmitter. On the other hand, hypercapnia acts indirectly by altering intracellular pH [19,20]. As a result, there is active H⁺ extrusion from the chemoreceptor cells in exchange for Na⁺. The rise in the intracellular Na⁺ concentration triggers the Na⁺-Ca²⁺ exchanger so that Na⁺ is in turn extruded in exchange for entry of Ca²⁺, causing the release of neurotransmitter.

The other possibility which may explain the lack of correlation between the two tests lies in the methodology of the single-breath CO₂ test. The single breath of 13% CO₂ may not have provided an adequate stimulus for the complete transduction of a ventilatory response. CO₂ is thought to activate the carotid body chemoreceptors by its intracellular acidifying capacity, as discussed above. The peripheral chemoreceptor response to CO₂ is therefore to a large extent indirectly dependent on the effects of hypercapnia on intracellular pH. Thus, fundamental differences in the methodology of the two tests may also explain our observation. As we excluded the ventilatory responses for 20 s after the stimulus breath of CO₂, it is unlikely that the responses seen in our study were influenced by central chemoreceptors. In studies that involve inactivation of peripheral chemoreceptors, there may be lag of up to 60 s before a ventilatory response is seen after a hypercapnic stimulus [4,7,21]. In our study, typical responses to hypoxia and single-breath CO₂ were seen within 20 s after the stimulus.

In conclusion, we have found that both the transient hypoxic and the single-breath CO₂ response tests are reproducible. In view of the lack of correlation between the two tests, however, we suggest that both should be undertaken in the assessment of peripheral chemoreflex since the presence of a hypoxic response does not imply a similar peripheral hypercapnic response and vice versa. The single-breath CO₂ test on its own is not an adequate test of the peripheral chemoreflex and cannot be used to predict hypoxic chemosensitivity. These tests may be separately and independently abnormal in certain disease states such as chronic heart failure and cyanotic congenital heart disease.

Limitations of study

With transient hypoxic testing, both arterial oxygen desaturation and the resultant ventilatory response are fleeting in nature. The measurement of arterial

oxygen desaturation and the use of a two-breath period to define maximal minute ventilation in such conditions, although necessary under the constraints of the method, may give rise to errors and hence potential study limitations. To minimize errors, we used a pulse oximeter with a fast response time and with documented accuracy of $< \pm 3\%$ compared with direct measurements of arterial blood samples [22,23]. Shaw *et al.* [6] have also previously shown that the transient hypoxic method, as used by us, can give as good an index of hypoxic chemosensitivity as the progressive method.

Some subjects felt an acid taste of the 13% CO₂, although none developed a cough. The acid taste may have affected ventilation and is a potential drawback of this method. However, the overall results of our single-breath CO₂ test compare favourably with previous data [7,24].

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